

Drug-induced liver injury: Interactions between drug properties and host factors

Minjun Chen^{1,†}, Ayako Suzuki^{2,3,†}, Jürgen Borlak⁴, Raúl J. Andrade^{5,6,*}, M Isabel Lucena^{5,5}

¹Division of Bioinformatics and Biostatistics, National Center for Toxicological Research, US Food and Drug Administration, Jefferson, AR, United States; ²Gastroenterology, Central Arkansas Veterans Healthcare System, Little Rock, AR, United States; ³Department of Medicine, University of Arkansas for Medical Sciences, Little Rock, AR, United States; ⁴Center of Pharmacology and Toxicology, Hannover Medical School, Hannover, Germany; ⁵Unidad de Gestión Clínica de Enfermedades Digestivas, Servicio de Farmacología Clínica, Instituto de Investigación Biomédica de Málaga (IBIMA), Hospital Universitario Virgen de la Victoria, Universidad de Málaga, Málaga, Spain; ⁶Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (CIBERehd), Madrid, Spain

Summary

Idiosyncratic drug-induced liver injury (DILI) is a common cause for drug withdrawal from the market and although infrequent, DILI can result in serious clinical outcomes including acute liver failure and the need for liver transplantation. Eliminating the iatrogenic "harm" caused by a therapeutic intent is a priority in patient care. However, identifying culprit drugs and individuals at risk for DILI remains challenging. Apart from genetic factors predisposing individuals at risk, the role of the drugs' physicochemical and toxicological properties and their interactions with host and environmental factors need to be considered. The influence of these factors on mechanisms involved in DILI is multi-layered. In this review, we summarize current knowledge on 1) drug properties associated with hepatotoxicity, 2) host

E-mail address: andrade@uma.es (R.J. Andrade).

These authors equally contributed to the manuscript.

Abbreviations: DILI, drug-induced liver injury; HLA, human leucocyte antigen; DAMPs, the damage associated molecular pattern molecules; ADMET, absorption, distribution, metabolism, excretion and toxicity; GST, glutathione-S-transferase; ROS, reactive oxygen species; JNK, c-Jun N-terminal kinase; Nrf-2, nuclear factor erythroid 2-related factor 2; Keap-1, Kelch-like ECH-associated protein 1; mtDNA, mitochondrial DNA; MPT, mitochondrial permeability transition; BSEP, bile salt export pump; ATP, adenosine triphosphate; P-gp, P-glycoprotein; MRP, multidrug resistance-associated protein; NAT2, N-acetyltransferase 2; CYP450, cytochrome P450; GSTM1, glutathione S-transferase Mu 1; GSTT1, glutathione S-transferase theta 1; NSAIDs, non-steroidal anti-inflammatory drugs; GSH, gluthatione; POLG, polymerase (DNA directed), gamma; FXR, farnesoid X receptor; LPS, lipopolysaccharides; MELD, Model for end-stage liver disease; PARP-1, Poly-(ADP-Ribose) Polymerase-1; NAFLD, non-alcoholic fatty liver disease; SOD2, superoxide dismutase 2; GPX1, glutathione peroxidase; NASH, nonalcoholic steatohepatitis; UDPGT, UDP-glucuronosyltransferase; NRTIS, nucleoside reverse transcriptase Inhibitors: PPAR γ . peroxisome proliferator-activated receptor gamma; APC, antigen-presenting cell; MHC, major histocompatibility complex.



Journal of Hepatology 2015 vol. 63 503-514

factors considered to modify an individuals' risk for DILI and clinical phenotypes, and 3) drug-host interactions. We aim at clarifying knowledge gaps needed to be filled in as to improve risk stratification in patient care. We therefore broadly discuss relevant areas of future research. Emerging insight will stimulate new investigational approaches to facilitate the discovery of clinical DILI risk modifiers in the context of disease complexity and associated interactions with drug properties, and hence will be able to move towards safety personalized medicine.

© 2015 European Association for the Study of the Liver. Published by Elsevier B.V. Open access under CC BY-NC-ND license.

Introduction

Drug-induced hepatotoxicity is one of the major concerns in medical practice. Although it is relatively uncommon, drug-induced liver injury (DILI) is the leading cause of acute liver failure in the US and a major reason for liver transplantation [1]. Many marketed drugs, herbs and dietary supplements have a potential to cause liver injury. In preclinical studies, about 50% of candidate compounds present hepatic effects at supra-therapeutic dose and face drug attrition [2]. DILI is also a major cause for drug failure in clinical trials and frequently results in regulatory actions and drug withdrawal [3,4].

The incidence of DILI in general populations is about 14–19 per 100,000 inhabitants [5,6], while frequency estimated in a healthcare system is around 30-33 per 100,000 persons [7]. The reported incidence and severity of DILI varies among drugs [6,7], suggesting that drug properties have a role in DILI risk determination. Conversely, drugs with DILI potential cause liver injury only in a small portion of patients indicating that host factors play a major role in DILI development.

DILI is classified into intrinsic vs. idiosyncratic liver injury, reflecting a dominant role of drug toxicity (dose-dependent) vs. host factors (no dose dependence) in liver injury. With a few exceptions (i.e., acetaminophen), most of DILI experienced in humans are considered idiosyncratic. However, inflammatory stress may influence the dose-response curve towards

Keywords: Drug-induced liver injury; Drug physicochemical properties; Host factors; Drug-host Interaction; Pharmacogenetics; Drug metabolism; Drug clearance; Clinical toxicology.

Received 6 February 2015: received in revised form 1 April 2015: accepted 7 April 2015

^{*} Corresponding author. Address: Departamento de Medicina, Facultad de Medicina, Boulevard Louis Pasteur 32, 29071 Malaga, Spain. Tel.: +34 952 134242; fax: +34 952 131511.

sensitization for toxicity at therapeutic doses, making the two DILI types less distinct [8]. Indeed, around 10% of acetaminophen-induced acute liver failure cases occurred at recommended dosage, suggesting host factors modify individual risks of acetaminophen liver injury [9,10]. Besides, drug dosage is a well-known determinant of idiosyncratic DILI [11,12]. Thus, the two entities may rather coincide in human DILI.

The current mechanistic understanding of DILI is depicted in Fig. 1. The key mechanisms in DILI are two-fold: 1) drug/metabolite exposure to a threshold level, determined by the dose and drug handling of the liver, and 2) the adaptive immune response or "alarm-signalling" by the damage associated molecular pattern molecules (DAMPs) [13]. Cellular damage occurs at an intricate balance between toxic drug exposure and defence mechanisms. Once cells are damaged, innate and adaptive responses kick-in and play a significant role in driving tissue inflammation and injury. The degree of local tissue inflammation and injury, in a balance with tissue repair, influences overall tissue damage and determines clinical outcome. Drug exposure and properties of administered drugs play primary roles at the initial stages of cellular damage while host factors drive 'host responses' to toxic insults with the induction of cellular repair programs.

This review will systematically update the current knowledge on drug properties associated with hepatotoxicity, discuss various host factors that may contribute to individuals' DILI risks and clinical phenotypes, and allude to potential drug-host interactions aiming at providing a structured conceptual framework to guide future empirical research in this challenging field.

Key points

- Individual risks and clinical phenotypes of DILI are likely determined by a multi-faceted interplay between drugs' physicochemical and toxicological properties, host factors and the interactions among them.
- Drug properties contributing to initial cell damage include surpassing a threshold dose, physicochemical characteristics such as lipophilicity, formation of reactive metabolites, induction of oxidative stress, mitochondrial hazard and inhibition of hepatic transporters.
- Age, gender, genetic factors, pubertal development, hormonal and nutritional status, pregnancy, co-medications, underlying conditions and the gut microbiome influence key mechanistic components of DILI which can be classified into four categories: drug handling, toxicological responses, inflammation and immune responses, and the balance of tissue damage and repair.
- Further investigations on drug-host interactions are needed to integrate the drug signature data with patient clinical data that would enable the discovery of clinical DILI risk modifiers and their interactions with drug properties as to move towards safety personalized medicine.
- Developing new investigational approaches, involving bioinformatics and computer science may enhance the transferability of information and facilitate inter-disciplinary research in the field.



Fig. 1. Current mechanistic understanding in the initiation and progression events relevant to idiosyncratic drug-induced liver injury. Two mechanistic cascades, (A) Sterile inflammation caused by drug-induced cytotoxicity vs. (B) Immune response via antigen presenting cells (APCs) and/or helper T-cells. Drugs/ reactive metabolites exert direct toxicity or form adducts leading to haptenization. Cells respond by activating adaptive pathways. Injured hepatocytes release "danger signals", such as the damage associated molecular patterns molecules (DAMPs) which favour the release of pro-inflammatory cytokines to induce a T/B-cell response against hepatocytes. The HLA associations discovered in GWAS suggest that the adaptive immune response is an upstream event. The innate immune system can either co-stimulate the adaptive immunity or modulate the degree of inflammation and regeneration.

Drug properties related to DILI risk in humans

Drugs within a therapeutic class differ regarding their hepatic liability, suggesting that physicochemical and toxicological drug properties affect DILI risk. Typical examples are thiazolidinediones, of which troglitazone was withdrawn from market due to fatal hepatotoxicity, while rosiglitazone and pioglitazone were less harmful to the liver. Among drug properties, factors contributing to initial cell damage include surpassing a threshold dose, physicochemical characteristics, reactive metabolites formation, oxidative stress, mitochondrial hazard and inhibition of hepatic transporters.

Threshold dose

Idiosyncratic DILI is considered dose-independent; most DILI cases occur at therapeutic dose in an individual despite being well tolerated in the general populations. However, in preclinical testing, hepatotoxicity is often predicted at high drug exposure leading to several stress responses in hepatocytes [14]. The conventional concept of dose independency is being challenged [15]. Uetrecht firstly suggested that idiosyncratic DILI was rarely observed with drugs given at daily doses of $\leq 10 \text{ mg}$ [16] and many drugs withdrawn from market or issued with a boxed warning (e.g. nimesulide, bosentan) due to hepatotoxicity, were prescribed at daily doses ≥ 50 or 100 mg [17,18]. Moreover, DILI patients in large cohorts from Spain and Iceland [6,19] and 81% of non-acetaminophen DILI patients undergoing liver transplantation in the United States received medications with daily doses of \geq 50 mg [1]. Therefore, a significant association between daily dose and poor DILI outcome (i.e. liver failure, transplantation and death) exists and was also found in a systematic survey based on pharmaceutical databases [11]. These evidences suggest that surpassing a threshold dose is associated with an increased risk of triggering liver injury among the treated patients. Daily dose alone is, however, inadequate to reliably predict DILI risk from individual drugs because a majority of compounds needs \geq 50 mg to achieve efficacy [21].

Lipophilicity

A drug's physicochemical property is known to affect cellular uptakes and ADMET (absorption, distribution, metabolism, excretion and toxicity). Chen et al. [12] explored the impact of lipophilicity in combination with daily dose and found oral medications at high daily doses ($\geq 100 \text{ mg}$) and a lipophilicity of logP \geq 3 to be significantly associated with severe DILI. Their study demonstrated that both factors could individually predict hepatotoxicity, while the "rule-of-two", which combines dose and lipophilicity, performs better than daily dose alone, thus increasing the positive predictive value from 85% to 96% while decreasing the negative predictive value from 55% to 39%. Higher lipophilicity could enhance DILI risk by facilitating drug uptake from blood into hepatocytes, which conditions hepatic metabolism and may result in a greater amount of reactive metabolites, subsequently leading to an interaction with mitochondrial membranes and hepatocanalicuar transport [13,22]. Besides lipophilicity, other physiochemical properties as molecular weight and total polar surface area associate with in vivo toxicological outcomes [23,24].

Formation of reactive metabolites

Several lines of evidence suggests that the formation of reactive metabolites play a central role in the pathogenesis of idiosyncratic DILI [25]. Reactive metabolites can covalently bind proteins to form drug-protein adducts that might trigger immune-mediated reactions or exert direct toxicity [26,27]. Cholestasis may also be a consequence of the canalicular secretion of reactive metabolites or disintegration of labile glutathione and/or glucuronide conjugates thereby damaging cholangiocytes or triggering an immune response. However, for a given drug, there is no clear-cut correlation between the potential to form reactive metabolites in experimental conditions and the actual incidence of hepatotoxicity in humans [28]. Obach et al. [29] measured the formation of reactive metabolites in vitro and found that metabolism-dependent covalent binding with liver microsomes cannot distinguish hepatotoxic and non-hepatotoxic drugs. Another experimental study tested approximately 100 Merck drug candidates and found no correlation between liver toxicity observed from in vivo animal studies and the extent of covalent binding [30]. Within a given drug class, specific chemical structures can render the compound distinctly hepatotoxic. For instance, ebroditine, an antiulcer drug pharmacologically related to famotidine, carries a bromobenzene ring which undergoes metabolic activation to reactive epoxides [31]. Likewise, temafloxacin and trovafloxacin share a unique difluorinated side chain that does not occur in other quinolones with much less hepatotoxicity [32].

Oxidative stress

Oxidative damage in the liver could be a consequence of cytosolic oxidant stress after drug metabolism or could arise from oxidant

JOURNAL OF HEPATOLOGY

stress directly generated in mitochondria and the subsequent inflammatory cell response by injured hepatocytes. Oxidative stress is caused by an imbalance of reactive oxygen species (ROS) formation (c-Jun N-terminal kinase, JNK) and its detoxification by antioxidant defence systems (Nrf2/Keap1) [33]. The balance of products of oxidative stress, protective cellular defence and cytokines modulating inflammation may be critical for DILI susceptibility, severity and extent of injury. Increased ROS can directly damage DNA, proteins, enzymes, and lipids in cells and tissues and induce immune-mediated liver damage. Some drugs (e.g. valproic acid) can induce enhanced generation of ROS by interrupting the homeostasis of mitochondria respiratory chain and triggering JNK signalling pathway, to subsequently activate mitochondrial permeability and death of hepatocytes [33]. Recent reports suggest drug-induced oxidative stress also significantly correlate with DILI risk. Xu et al. identified ROS generation along with mitochondrial damage and intracellular glutathione depletion, as most important indicators contributing to hepatotoxicity as determined by high content imaging in primary human hepatocyte cultures [34].

Mitochondrial liability

Mitochondrial dysfunction plays a critical role in the pathogenesis of DILI by alteration of metabolic pathways and damage to mitochondrial components [33,35]. Drugs such as stavudine and amiodarone can induce steatosis/steatohepatitis by severely altering mitochondrial function. Mitochondrial damage could trigger hepatic necrosis and/or apoptosis, leading to activation of cell death signalling pathways such as JNK when a critical mitochondrial death threshold is surpassed [35,36]. This view challenges the traditional paradigm, indicating that cell death is rather an active process involving mitochondria thereby determining the fate of cells as opposed to overwhelming biochemical injury [36]. Specifically, drugs can impair mitochondrial respiration (valproic acid) and/or β-oxidation (aspirin, tamoxifen), trigger mitochondrial membrane disruption (diclofenac) and damage mtDNA (tacrine) [37-39]. Interestingly, Porceddu et al. [40] reported a significant association between loss of mitochondrial integrity and the potential to cause DILI, based on the analysis of 124 chemicals/drugs.

Inhibition of BSEP and other hepatobiliary transporters

Hepatobiliary transporters, and particularly the canalicular adenosine triphosphate (ATP)-dependent bile salt export pump (BSEP), are responsible for the biliary excretion of several organic compounds including bile acids. An impaired function of BSEP determines the accumulation of cytotoxic bile acids in hepatocytes leading to the induction of oxidative stress and/or apoptosis and necrosis by FAS-mediated pathways [41]. Drugs and/or metabolites with capacity to inhibit BSEP in vitro have potential to cause DILI as has been shown by Morgan et al. using BSEP-inverted vesicles [42]. Although this approach enables preclinical drug testing with some drugs shown to be potent BSEP inhibitors and have either been withdrawn from the market (troglitazone) or received warnings (imatinib) for hepatotoxicity, others (pioglitazone, simvastatin) have a low potential for DILI risk. Hence, BSEP inhibitory potency alone is insufficient for determining DILI risk and additional factors should be considered. Recently, Aleo et al. demonstrated that drugs which carry

a more serious DILI risk influence both BSEP and mitochondrial activities [43]. Mitochondrial dysfunction would result in impaired ATP production, and in conjunction with BSEP inhibition, might explain the synergistic link between mitochondria and ATP-dependent transporters such as BSEP in DILI.

The hepatic canalicular transporter P-glycoprotein (P-gp) is a well-known determinant in multidrug resistance in chemotherapy [44]. Other hepatobiliary transporters of the multidrug resistance protein (MRP) family are also involved in the excretion of conjugated organic anions, bilirubin and drug metabolites. Recent studies suggest that the consideration of MRP2/3/4 inhibition could improve the correlation with DILI risk in humans as compared with BSEP inhibition alone [45], suggesting that defects in transporters function modify drug disposition. Owing to the fact that hepatocytes are highly polarized and transporters function either bi- or mono-directional, the host and drug interactions may lead to different phenotypes of DILI (i.e. cholestasis, hepatocellular, steatosis).

Host factors modifying DILI risks and clinical phenotypes

Host factors contributing to individual susceptibility and clinical phenotypes of DILI have not been systematically investigated. In this section, we will provide cross-disciplinary view over host factors influencing key mechanistic components of DILI, classified into four categories: drug handling, toxicological responses, inflammation and immune responses, and imbalance of tissue damage and induction of repair processes.

Host factors influencing drug handling

Factors that modify the level of exposure to the reactive metabolites and/or alter the disposition of the drug may critically influence the development of DILI. In individual cases, drug therapy adjustments appear to change a drug's hepatotoxic potential; e.g. reducing the dose of mianserin [46] and prolonged dose intervals of gefitinib [47] eliminated risk of hepatotoxicity while atorvastatin dose escalation increased the risk of hepatotoxicity [20]. These observations underpin the need of surpassing a threshold dose to induce DILI in a unique susceptible individual [20]. Inter-individual differences in drug tissue concentration are further influenced by oral bioavailability, volume of distribution, visceral blood flow, drug metabolism, nutritional status, excretion/transport, age and genetic and epigenetic factors.

Aging is known to influence the pharmacokinetics of drugs due to decreased renal function and cytochrome-mediated hepatic metabolism, while reduced conjugation reactions seem to be restricted to older frail patients [48]. Hence, older age likely enhances DILI susceptibility. This concept, however, has not been supported by data from large national DILI registries. In the Spanish DILI Registry 46% of DILI patients were ≥60 years of age and the US Drug-Induced Liver Injury Network (DILIN) reported 18.5% of DILI patients to be 65 years or older [49,50]. In a population-based study done in Iceland, a relationship between DILI incidence and increasing age was observed, probably related to a greater exposure to polypharmacy in older subjects [6]. Apparently, the type of liver injury differed with age with younger patients presenting more frequently hepatocellular damage as compared to cholestatic/mixed injury seen in the old [49,51]. The risk of developing valproic acid-induced

hepatotoxicity with fatal outcomes is higher in children below the age of two [52]. Hepatotoxicity induced by isoniazid appears to be more frequent in older patients. A retrospective study in 3377 adults receiving isoniazid therapy demonstrated that the DILI incidence was about two-fold amongst 35–49 years old and almost five-fold in \geq 50 years old patients as compared to the 25–34 years old ones [53].

Furthermore, gene expression of drug metabolizing enzymes and transporters vary significantly among individuals, being influenced by genetic variants, epigenetic alterations, age, gender, hormones, nutrition, alcohol, and co-medications [54]. Genetic polymorphisms of drug metabolizing enzymes are estimated to influence the clinical outcome in 20–25% of all drug therapies [54]. Some racial differences in DILI caused by anti-tuberculosis drugs have been attributed to variants of drug metabolizing genes coding for NAT2, CYP2E1, GSTM1 and GSTT1 [55]. Thus, polymorphisms of drug metabolizing enzymes and transporters are considered as one of the key contributors in an individual's DILI risk [56].

Gender, pubertal development, sex hormones, pregnancy and growth hormone levels also influence drug metabolizing enzymes [57]. For instance, men have a higher clearance rate of acetaminophen than women due to higher glucuronidation rates, while CYP3A4, a major drug metabolizing enzyme, is expressed at a higher rate in women [58]. Furthermore, cytokines released in systemic infection inflammation significantly represses activities of cytochrome P450 monooxygenases and transporters [59,60]. Consequently, in patients with systemic inflammatory response syndrome, detoxification processes may significantly decrease possibly requiring dose adjustment.

Lifestyle, disease conditions, and co-medications also modify individual's drug handling capability. Alcohol and high fat diets are known to induce CYPs 2E1 and 4A. Alcohol-induced increase in CYP2E1 has been associated with an increased risk of acetaminophen-induced liver injury in humans, which is explained by an increased generation of the reactive metabolite N-acetyl-p-benzoquinone imine (NAPQI) [61]. Malnutrition and cellular senescence could result in decreased xenobiotic clearance and subsequently lead to slower drug elimination and higher drug plasma levels. Additionally, several marketed drugs are known to inhibit/induce specific drug metabolizing enzymes and transporters [62], which potentially alter reactive metabolite formation, drug conjugation, and/or drug elimination, and therefore modifying an individual's DILI risk [61,62].

Host factors modifying toxicological responses

Drugs initiate cellular damage through diverse mechanisms: reactive metabolite formation, which leads to covalent binding to cellular proteins, oxidative stress, endoplasmic reticulum stress, mitochondrial injury, DNA damage, epigenetic modifications, and/or inhibition of bile acid excretion (Fig. 1). Various patients' host factors may influence toxicological responses and modify the risks of developing cellular damage.

Specifically, risk of inducing cellular damage through reactive metabolites is affected by cellular detoxification mechanisms. Patients with genetic defects in GST were reported to have an increased risk of developing DILI caused by anti-tuberculosis drugs [63], NSAIDs and antibacterials [64]. Slow acetylators of NAT2 were also associated with moderate to severe DILI related to anti-tuberculosis drugs [65]. Thus, at a given amount of

JOURNAL OF HEPATOLOGY

reactive metabolite formation, those with diminished cellular detoxification are at a higher risk of developing DILI.

Induction of cellular oxidative stress is another major toxicological insult caused by drugs. The degree of drug-induced oxidative insult may be modified by host's pre-existing increased cellular oxidants, increased substrates for oxidative reactions (e.g., steatosis, lipid peroxidation), and/or decreased anti-oxidants. Patients with functional polymorphisms in mitochondrial superoxide dismutase and glutathione peroxidase have a higher risk of developing DILI, especially for those culprit drugs that are hazardous for mitochondria and/or form highly reactive intermediates [63,66]. Other host factors influencing cellular oxidative stress are listed in Table 1 [26,67,68]. A female-specific susceptibility to oxidative stress in idiosyncratic DILI has been reported [49].

Host factors influencing mitochondrial functions are listed in Table 1 [26,69,70]. In normal mitochondrial biology, significant amount of ROS is produced and usually appropriately detoxified

Mechanistic factors	Drug properties		Host responses		
	Specific factors	Examples	Specific factors	Examples	
Threshold dose	Daily dose [11,12]	Duloxetine, gefitinib, bosentan, tacrine, leflunomide, methotrexate	Drug absorption and hepatic delivery	Gastric emptying, gastrointestinal transit, nutrition, aging, atherosclerosis, portal hypertension	
	Bioavailability⁺[12]	Vancomycin, aminoglycosides, rifaximin, cromoglicate			
	Long half-life	Azithromycin, tamoxifen	A reduced drug clearance (i.e., prolongs half-life)	High body fat, elderly, renal dysfunction, hepatic dysfunction	
Covalent binding	Significant hepatic metabolism [119]	Atorvastatin, tacrolimus, disulfiram, terbinafine	Hepatic drug metabolism	Genotypes of drug metabolizing enzymes, age [120], sex [57], Inducers/inhibitors of drug metabolizing enzymes (e.g., co- medications, alcohol, and diets)	
	Reactive metabolite generation [38]	Acetaminophen, trovafloxacin⁺, isoniazid, phenytoin, carbamazepine, valproic acid, diclofenac	Impaired cellular proteins, repair/ degradation [121,122]	Reduction of thioredoxin/thioredoxin reductase, glutathione reductase, methionine sulfoxide reductase	
Oxidative stress	Increase intracellular (e.g. mitochondria) oxidants [33]	Acetaminophen, troglitazone⁺, flutamide, nimesulide⁺, valproic acid, diclofenac	Increase cellular oxidants	Obesity/insulin resistance/NAFLD, advanced cellular senescence	
			Increase lipid peroxidation	Fatty liver	
			Depletion of antioxidants	Aging, obesity/insulin resistance/ NAFLD, genotypes related to cellular antioxidation (e.g., SOD2, GPx1) [26], nutrition, lack of estrogens [123]	
Mitochondrial liability	Impair mitochondrial respiration [38]	Paroxetine, valproic acid, troglitazone ⁺ , nefazodone+	Mitochondrial dysfunction	Genetic variants of mitochondrial enzymes, age, sex, sex hormones,	
	Inhibit beta-oxidation [38]	Amineptine⁺, ibuprofen, valproic acid, minocycline, aspirin		advanced cellular senescence (e.g., insulin resistance/NASH, chronic inflammation)	
	Trigger mitochondrial membrane disruption [26]	Ciprofloxacin, diclofenac, indomethacin			
	Damage mitochondrial mtDNA [26]	Tacrine, tamoxifen, stavudine and other NRTIs	Impair mitochondrial DNA repair	Genotypes of mitochondrial DNA polymerase γ [73]	
Hepatic transporters inhibition	Inhibit BSEP [42]	Troglitazone ⁺ , bosentan, erythromycin, estradiol, simvastatin, rifampin, imatinib, nefazodone+	Hepatic transporter regulations	Genotypes related to transporters (e.g., BSEP, MRP2/3/4), co- medications, release of bacterial endotoxin due to increased intestinal permeability, altered hepatic FXR (e.g., NASH [124], bile acid pool and components [125])	
	Inhibit other hepatic transporters (e.g., MDR3/ MPR2/MPR3/ MPR4) [42]	Itraconazole (MDR3), zafirlukast (MRP2), atorvastatin (MRP3/4), indomethacin (MRP3/4)	Impair energy supply for hepatic transporters	Aging, cellular senescence/ mitochondrial dysfunction	

Table 1. Overview of drug/host factors influencing specific mechanisms involved in idiosyncratic drug-induced liver injury.

(*Continued on next page*)



Table 1 (Continued)

Mechanistic factors	Drug properties		Host responses	
	Specific factors	Examples	Specific factors	Examples
Inflammation and immune responses	Anti-inflammatory drugs	Aspirin [126], coxibs [127], statins [128]	Pro-inflammatory conditions	Increased influx of LPS (e.g., alcohol abuse, intestinal diseases) [86,87], altered microbiome [129], chronic inflammatory diseases and viral infections, obesity [130], progesterone [81], depletion of bile acid pool [131]
			Anti-inflammatory conditions	Estrogens [81], androgens [26], co-medications (anti-inflammatory drugs)
	Anti-TNFα drugs and other biological products	Azathioprine, leflunomide, tacrolimus, adalimumab, infliximab	Modify immune responses	HLA, sex [27], sex hormones [132], co-medications (e.g., immunosuppressant, immunomodulator), epigenetic alterations (e.g., hydralazine and procainamide)[133], gut microbiota [129]
	Immunosuppressants and immunomodulators	Glucocorticoids, opioids [134], antihistamines [135], statins [136]		
Tissue injury and repair	Dominant induction of necrosis <i>vs.</i> apoptosis	Acetaminophen, troglitazone⁺, flutamide, diclofenac	Apoptosis <i>vs.</i> necrosis	Sex [106], sex hormones [106], cellular energy supply [137]
	Impair tissue repair	Hydralazine derivatives (histone acetylation inhibition) [97], sympathetic stimulants [138, 139]	Tissue repair	Aging [103], advanced cellular senescence [103], co-medications [93], altered FXR [140], sex [141], sex hormones [142]

^{*}Drugs of very low bioavailability were associated with few DILI reports (e.g., acarbose).

*Drugs that were withdrawn from markets worldwidely or in some countries.

[71]. However, mitochondrial aging, partly due to accumulated oxidative mitochondrial DNA damage [38], may be affected by other host factors such as over-nutrition (e.g., obesity, insulin resistance, diabetes, and NASH) and alcohol [38,72]. Damaged mitochondrial DNA is repaired and maintained by mitochondrial DNA polymerase γ , encoded by the nuclear gene *POLG*. A recent gene-association study showed that about 50% of cases with valproate-induced liver injury were heterozygous for *POLG* substitution mutations and its odds ratio was estimated as high as 24 [73]. Individuals with carnitine deficiency were also associated with an increased risk of valproate-induced liver injury [74] while carnitine appears to be protective against valproate-induced liver injury and improve survival in severe cases.

Inhibition of bile acid transporter leads to intrahepatocellular bile acid accumulation while inhibition of phosphatidyl choline excretion (MDR2/3) alters bile composition and leads to cholangiocyte injury [75]. As shown in Table 1, hepatic transporters are influenced by genetic variations, co-medications, bacterial endotoxins and the farconoid xenosensing receptor (FXR), which functions as a bile acid sensor and acts as a key regulator of metabolic processes [41].

Bile acids salts are anionic detergents and highly toxic to the cells. In bile, mixed micelle formation with cholesterol, phospholipids, bile pigments, proteins, and inorganic electrolytes protects cholangiocytes from the toxic detergent effect of bile acid salts. Dysfunction of MDR3/ABCB4 (phosphatidyl choline translocation across canaliculus membranes, regulated by FXR) has been associated with clinical cholestasis, presumably via inhibition of micelle formation, releasing free bile acids salts in bile [76]. Patients with primary biliary cirrhosis and extrahepatic bile obstruction have decreased biliary bicarbonate secretion measured by positron emission tomography [77,78], suggesting a potential susceptibility to drugs influencing bile components (i.e., itraconazole) [78].

Host factors modulating inflammation and immune responses

Innate/adaptive immune response plays a key role in inducing inflammation and determining the degree of 'injury' (Fig. 1). Host factors known to modulate inflammation and immune response which, in turn, may influence DILI susceptibility will be discussed below.

Several genetic variants in the HLA regions were identified as risk factors for DILI [56]. Carriers of the HLA-B*57:01 genotype are at an 80-fold increased risk of flucloxacillin-induced DILI [79]. DILI caused by other drugs (e.g. lumiracoxib, lapatanib, ticlopidine, amoxicillin-clavulanate and ximelagatran) are also associated with HLA genotypes [21]. Even causal drugs not accompanied by hypersensitivity features show the association with the HLA haplotypes, suggesting an important role of the immune system in DILI [21].

Gender and sex hormones are well-known to influence inflammation and immune response. An immune-mediated DILI model showed gender bias in immune response and inflammation; more severe hepatitis, more antibody production, and a higher level of pro-inflammatory hepatic cytokines in females *vs.* males [80]. Indeed, females with DILI are at a higher risk of developing acute liver failure or requiring liver transplantation [19,49]. In halothane-induced DILI, estrogens reduce liver injury in mice while progesterone exacerbates the damage possibly by modulating inflammation and immune response. Indeed, increased hepatic neutrophils and up-regulated hepatic mRNA levels of pro-inflammatory cytokines were noted with

Review

Racial differences in inflammation and immune response are also known. African-Americans are at a higher risk of developing chronic DILI (defined as persistent liver alteration beyond six months of DILI recognition), while Asians are associated with earlier development of liver-related death or liver transplantation [82]. Potential race-associated genetic variants enhancing inflammation or adaptive immune response are warranted future investigations.

Immune and inflammatory responses are also influenced by medications co-administered at the time of drug exposure. Previous data-mining using a large spontaneous adverse event reporting system discovered latent associations between reduced reporting frequency of liver events and various co-reported medications. Among the identified medications, anti-inflammatory agents and immunosuppressants were disproportionally prevalent [83,84]. Despite the preliminary nature of these observations, the associations suggest that the concomitant use of anti-inflammatory and immunosuppressant agents may modulate host immune response and inflammation and impact DILI occurrence. Other host factors potentially influencing inflammation and immune response are listed in Table 1.

The gut–liver axis plays a role in DILI. Increased intestinal permeability due to damaged intestinal mucosal barrier increases hepatic endotoxin influx, which in turn activates Kupffer cells and the production of pro-inflammatory cytokines, arachidonic acid metabolites and ROS in the liver [85]. In experimental models, intestine-derived endotoxin or co-administration of LPS enhances liver injury induced by chemicals [86,87], while decreased intestinal permeability reduced liver injury [88]. Likely, a disrupted mucosal barrier induced by drugs (e.g. NSAID), alcohol abuse, or intestinal disorders as seen with celiac disease and inflammatory bowel disease, or acute enterocolitis can act synergistically enhancing liver damage caused by hepatotoxic drugs [14].

Whether pre-existing chronic liver diseases enhances the risk of hepatotoxicity is hampered by the fact that recrudescence of inflammation can go undistinguished from true injury induced by a drug. A few examples, however, suggest potential enhancement of drug hepatotoxicity by existing chronic inflammation (or chronic viral infection). A previous retrospective study showed that patients with pre-existing chronic liver injury are at an increased risk of acute liver injury following acetaminophen overdose [89]. Severe DILI cases caused by anti-retroviral medications are more commonly observed among patients co-infected with hepatitis B and/or C virus [90]. Further, chronic hepatitis C virus infection, human immunodeficiency virus (HIV) infection, and autoimmune disease were associated with an increased risk of DILI caused by anti-tuberculosis drug therapy [91,92].

Host factors modifying cell death, tissue injury and repair

The balance between tissue injury and repair needs to be considered with impaired tissue repair worsening the condition leading to poor clinical outcome. This concept is supported by clinical studies, where the impact of co-medications on DILI outcome in patients with acetaminophen-associated liver injury was examined [93,94]. Briefly, co-medications with drugs which ameliorate liver injury and/or enhance liver repair in animal experiments (e.g., statins, fibrates, β -blockers, NSAIDs) were associated with a

decreased likelihood of fatality (or lower MELD scores) among acetaminophen-associated liver injury while co-medications with drugs enhancing liver injury and/or impairing liver regeneration (i.e., sympathetic stimulants) were associated with an increased likelihood of fatality [93,94]. Potential beneficial impacts of lipid lowering drugs (i.e., statins, fibrates) and anti-inflammatory agents (e.g., NSAIDs, immunosuppressants) are associated with improved clinical outcomes in patients diagnosed with dyslipidemia and collagen diseases among DILI cases [94,95].

JOURNAL OF HEPATOLOGY

Epigenetic modifications of host chromatin may impair regeneration following injury [96,97]. Loss of histone acetylation results in impaired liver regeneration in mice after toxic injury [96]. Impaired histone acetylation induced by todralazine (a hydralazine derivative) also results in impaired liver regeneration, which was correlated with clinical cases of drug-induced acute liver failure [97]. Additionally, nutritional deficiencies cause epigenetic modifications, which potentially alter individual susceptibility to hepatotoxicity. Deficiencies of folic acids, vitamin B12, and choline induce methyl donor depletion, contributing to hypomethylation of genes in cellular metabolism and hepatocyte differentiation [98-100]. Folic acid deficiency is associated with more severe liver damage in ethanol-fed micropigs [101.102] while folic acid supplementation has been associated with a reduced reporting frequency of liver events across different agents with hepatotoxic potential in previous data-mining analyses [83,84].

Age-related decline of mitochondrial function may also compromise energy supply for cellular metabolism and tissue regeneration [71,103]. In patients with hepatitis A, a likelihood of poor clinical outcomes increases with increased age [104]. Decompensated cirrhosis is another factor of poor outcome. Such patients require specific care in the selection of medications, and drugs with significant hepatic metabolism should be avoided [105].

Toxic insults can induce different forms for cell death. Unlike apoptosis, necrotic cell death leads to plasma membrane disturbance and subsequent releases of its cellular contents, which may induce an inflammatory response. Sexual dimorphism was observed in such cell death regulations in other systems [106,107]. An immune-mediated nephritis mouse model evidenced more apoptosis in females but more necrosis in males. The observed gender-biased in cell death was partially mediated by estrogen and Poly-(ADP-Ribose) Polymerase-1 (PARP-1) [106]. In one recent clinical analysis of DILI cases, the frequency of apoptosis was increased in women at a given injury pattern [108]. Further investigations are warranted to delineate the suspected sex difference in cell death and its clinical relevance.

Drug-host interaction: what do we know and what should we know, and how should we approach it

Both drug properties and host factors are multi-layered, influencing multiple mechanisms, and likely interact at multiple levels to determine DILI susceptibility, clinical phenotypes and outcome. Table 1 provides a structured summary of drug properties and host factors relevant to human DILI, which is organized based on mechanistic elements. Some combinations of drugs and host factors may exert additive interactions on DILI risks, which may explain clinical observations of high-risk populations for specific agents. A few examples with suggested mechanisms are provided in Table 2. A previous data-mining analysis showed that

Table 2. Specific drug-host interactions influencing risks of idiosyncratic drug-induced liver injury [14].

Causative DILI agent	Drug properties	Host resp	Possible consequence of	
		Known risk factors	Suggested mechanisms	drug-host interaction
Valproic acid	High solubility, extensive metabolism	 Young age (in particular <2-3 years) Antiepileptic co-medication 	 Different CYP2C9 enzyme activity among developmental stages in children Enhance 4-ene-valproic acid metabolite formation by inducing CYP activities (CYP2A6, CYP2C9) 	Enhanced reactive metabolite generation
	Mitochondrial liability	 Metabolic defects (impaired hepatic mitochondrial functions) Genetic variations in <i>POLG</i> (mitochondrial DNA polymerase γ) 	 Valproic acid undergoes β-oxidation and competes with endogenous lipids for enzymes and the mitochondrial CoA pool in this pathway. Impaired mitochondrial DNA replication 	Mitochondrial damage
Atorvastatin	High lipophilicity	• Older age	Reduction in drug clearance	Threshold dose
	Extensively metabolized by CYP3A4 Reactive metabolites	 Genotypes of drug metabolizing enzymes Co-medications (e.g. ketoconazole, nefazodone, ritonavir, erythromycin) 	Functional CYP3A4 polymorphisms Inducers/inhibitors of CYP3A4	Enhanced reactive metabolite generation
	Immunomodulation	Women, older age	Autoimmune phenotype	 Autoimmune hepatitis triggered by statins
Diclofenac	 High solubility Extensive metabolism Enterohepatic circulation 	 Genotypes of drug metabolizing enzymes and transporters Co-medications 	 Underlying genetic polymorphisms in drug metabolizing enzymes (<i>CYP2C8</i>, <i>UDPGT 2B7</i>, <i>GST</i>), and hepato- canalicular transporters (<i>BSEP</i>, <i>MRP2</i>, <i>MRP4</i>) Inducers/inhibitors of drug metabolizing enzymes and transporters. 	 Enhanced reactive metabolite generation and/or Delayed clearance of drug/ metabolites in hepatocytes, increase hepatic exposure
	 Formation of acyl glucuronide and oxidative electrophilic quinine imines metabolites 	Genotypes of anti-oxidant system	Polymorphisms of SOD2 and GPX1	 Impaired anti-oxidation
	Mitochondrial liability	Preexisting diseases: osteoarthritis, rheumatoid arthritis, viral infections, diabetes mellitus	 Pre-existing mitochondrial dysfunction: Electrophiles derived from reactive metabolites causing mitochondrial dysfunction 	Mitochondrial damage
	Interaction with APC via MHC type II molecule	• HLA genotype [PPARy-associated SNP, rs17036170: OR(95%CI) of 11.3(4.9-25.9)]	Innate and adaptive immune mediated	Enhanced immune response
	Intestinal toxicity	Gut microbiome Pre-existing chronic inflammatory conditions	 Increased LPS influx due to compromised mucosal barrier, induced by diclofenac Modulation of hepatic inflammation via C-reactive protein 	Enhanced hepatic inflammation
Amoxicillin clavulanate	 High solubility Multi-drug regimens Poor metabolism Biliary excretion 	• Older men (>65 years)	Impaired drug clearance and prolonged exposure of the bile duct cells to the drug metabolite through canalicular excretion	Predominant cholestatic/ mixed injury among older subjects
	Interaction with APC via MHC type II molecule	Repeated prescription HLA genotypes: North Europe, DRB1*1501-DRB1*0602 and HLA-A*0201; Spanish, hepatocellular injury: HLA-A*3002 (OR = 6.7) and HLA-B*1801 (OR = 2.9), cholestatic injury: DRB1*1501-DRB1*0602 Racial disparities: Northern vs. Southern Europeans Caucasian	Innate and adaptive Immune mediated	• Enhanced immune response

mitochondrial liability was more prevalent among the drugs with an increased pediatric reporting frequency, while cholestatic manifestation, high lipophilicity and biliary excretion were more common among the drugs associated with a higher reporting frequency in the elderly, which might be explained by interactions between specific drug properties and age-biased attributes [51]. Drug-host interactions also appear to exist between specific drug properties and host genetic variants. Lucena et al. found that SOD2Ala/Ala genotype was associated with an increased risk of developing cholestatic/mixed injury induced by drugs with mitochondrial hazard [66]. Ulzurrun et al. suggested positive interaction between drugs containing a carbocyclic system with aromatic rings (e.g. NSAIDs) and a genetic variant, ABCC11 c.133 CC in DILI susceptibility [109]. Lastly, sexual dimorphism (XX vs. XY) may contribute gender-specific susceptibility of neurons and splenocytes to different cytotoxic agents, suggesting gender bias in cellular toxicological responses [110]. Whether hepatocytes or cholangiocytes exerts similar gender-biased toxicological responses requires future investigation.

Collectively, a conceptual framework explaining the relevance of drug-host interactions in human DILI is depicted in Fig. 2. The proverb of "the blind men and the elephant" teaches us the manifold nature of truth; in the story, every one of the blind men touches different parts of the elephant and describes it differently without knowing that all stems from the same animal. Through this analogy, we intent to highlight the different mechanisms underlying human DILI. Future investigations targeting drug-host interactions in an integrative system analysis will favour unravelling the determinants that overlap and potentiate each other on DILI. In this regard, recent progress in differentiating induced pluripotent stem cells makes it possible to develop



Fig. 2. Conceptual framework explaining drug-host interactions in human DILI. Two key players in DILI, drug and host factors may interact in a multifaceted manner at different functional pathways and determine individual susceptibility, clinical phenotype and outcome. Mechanisms involved in the initiation of cellular injury are likely drug specific and may occur as consequence of the interaction between specific drug properties and host-specific activities. Once injury is established host responses to the injury insult (i.e., immune response, inflammation, tissue injury and repair) are mainly determined by host factors. Such responses are likely modulated by various host factors such as age, gender, genetic factors, lifestyles, disease conditions and co-medications.

JOURNAL OF HEPATOLOGY

patient-specific hepatocytes as a "host dependent" assay system to investigate drug-host interactions [111]. On the other hand, introducing advanced bioinformatics methodologies, machine learning [112], topic modelling [113], network analysis [114] and deep learning techniques [115] to clinical analysis will unmask hidden patterns/associations. Inter-disciplinary translation integrating preclinical knowledge, drug properties and clinical phenotype is of critical importance for a better understanding of human DILI. Development of standardized nomenclature, electronic form of knowledge base for hepatotoxic drugs and drug properties [116], ranking/classification of post-marketing safety profiles [117], and bioinformatics infrastructure to support discovery-driven research will enhance the transferability of information and facilitate inter-disciplinary research in the field.

Perspectives

This review aimed at highlighting current knowledge on drug properties, host factors and drug-host interactions in human DILI and identifying knowledge gaps to stimulate future investigation. As individual risks and clinical phenotypes of DILI are likely determined by a multi-faceted interaction between drug properties and host factors, a new paradigm of DILI studies should be directed to address not only host factors or drug properties alone but their interactions. Developing new investigational approaches involving bioinformatics and computer science may become crucial in such future investigations. Indeed, preclinical safety assessment is currently based on the paradigm "high doses in healthy animals". However, biological responses to drug treatment will inevitably differ in disease. Therefore, the utility of experimental models that simulate host conditions should be considered [118].

Current knowledge is still limited and insufficient for accurate DILI risk prediction. Further investigations targeting drug-host interactions will enable establishing patient's risk stratification and the development of a safety personalized medicine.

Financial support

This study was supported by the research grant P10-CTS-6470, PI12-00620, PI12-00378 and the Agencia Española del Medicamento y Productos Sanitarios (AEMPS). CIBERehd is funded by Instituto de Salud Carlos III.

Author JB gratefully acknowledges support from The Virtual Liver Network (grant 031 6154) of the German Federal Ministry of Education and Research (BMBF). Part of this work was also funded by the Lower Saxony Ministry of Culture and Sciences and the Volkswagen Foundation, Germany to JB. Grant number: 25A.5-7251-99-3/00.

Conflict of interest

The authors disclose the following: The views presented in this article do not necessarily reflect those of the U.S. Food and Drug Administration.

The authors who have taken part in this study declared that they do not have anything to disclose regarding funding or conflict of interest with respect to this manuscript.

Acknowledgement

We appreciated Drs. Weida Tong, John Senior, and Mark Avigan for their comments and/or discussions. We also thank the discussions amongst the Liver Toxicity Knowledge Base (LTKB) interest group.

References

- Reuben A, Koch DG, Lee WM. Drug-induced acute liver failure: results of a US multicenter, prospective study. Hepatology 2010;52:2065–2076.
- [2] Amacher DE. Serum transaminase elevations as indicators of hepatic injury following the administration of drugs. Regul Toxicol Pharmacol 1998;27:119–130.
- [3] Watkins PB. Drug safety sciences and the bottleneck in drug development. Clin Pharmacol Ther 2011;89:788–790.
- [4] Chen M, Zhang J, Wang Y, Liu Z, Kelly R, Zhou G, et al. Liver Toxicity Knowledge Base (LTKB) – A systems approach to a complex endpoint. Clin Pharmacol Ther 2013;95:409–412.
- [5] Sgro C, Clinard F, Ouazir K, Chanay H, Allard C, Guilleminet C, et al. Incidence of drug-induced hepatic injuries: a French population-based study. Hepatology 2002;36:451–455.
- [6] Bjornsson ES, Bergmann OM, Bjornsson HK, Kvaran RB, Olafsson S. Incidence, presentation, and outcomes in patients with drug-induced liver injury in the general population of Iceland. Gastroenterology 2013;144:e1413.
- [7] Shin J, Hunt CM, Suzuki A, Papay JI, Beach KJ, Cheetham TC. Characterizing phenotypes and outcomes of drug-associated liver injury using electronic medical record data. Pharmacoepidemiol Drug Saf 2013;22:190–198.
- [8] Roth RA, Ganey PE. Intrinsic versus idiosyncratic drug-induced hepatotoxicity – Two villains or one? J Pharmacol Exp Ther 2010;332:692–697.
- [9] Larson AM, Polson J, Fontana RJ, Davern TJ, Lalani E, Hynan LS, et al. Acetaminophen-induced acute liver failure: results of a United States multicenter, prospective study. Hepatology 2005;42:1364–1372.
- [10] Gulmez SE, Larrey D, Pageaux GP, Lignot S, Lassalle R, Jove J, et al. Transplantation for acute liver failure in patients exposed to NSAIDs or paracetamol (acetaminophen): the multinational case-population SALT study. Drug Saf 2013;36:135–144.
- [11] Lammert C, Einarsson S, Saha C, Niklasson A, Bjornsson E, Chalasani N. Relationship between daily dose of oral medications and idiosyncratic drug-induced liver injury: search for signals. Hepatology 2008;47:2003–2009.
- [12] Chen M, Borlak J, Tong W. High lipophilicity and high daily dose of oral medications are associated with significant risk for drug-induced liver injury. Hepatology 2013;58:388–396.
- [13] Kaplowitz N. Avoiding idiosyncratic DILI: two is better than one. Hepatology 2013;58:15–17.
- [14] Kaplowitz N, DeLeve LD. Drug-induced liver disease. 3rd ed. Waltham, MA, USA: Academic Press; 2013.
- [15] Senior JR. What is idiosyncratic hepatotoxicity? What is it not? Hepatology 2008;47:1813–1815.
- [16] Uetrecht JP. New concepts in immunology relevant to idiosyncratic drug reactions: the "danger hypothesis" and innate immune system. Chem Res Toxicol 1999;12:387–395.
- [17] Walgren JL, Mitchell MD, Thompson DC. Role of metabolism in druginduced idiosyncratic hepatotoxicity. Crit Rev Toxicol 2005;35:325–361.
- [18] Stepan AF, Walker DP, Bauman J, Price DA, Baillie TA, Kalgutkar AS, et al. Structural alert/reactive metabolite concept as applied in medicinal chemistry to mitigate the risk of idiosyncratic drug toxicity: a perspective based on the critical examination of trends in the top 200 drugs marketed in the United States. Chem Res Toxicol 2011;24:1345–1410.
- [19] Andrade RJ, Lucena MI, Fernández MC, Pelaez G, Pachkoria K, García-Ruiz E, et al. Drug-induced liver injury: an analysis of 461 incidences submitted to the Spanish registry over a 10-year period. Gastroenterology 2005;129:512–521.
- [20] Carrascosa MF, Salcines-Caviedes JR, Lucena M Isabel, Andrade RJ. Acute liver failure following atorvastatin dose escalation: is there a threshold dose for idiosyncratic hepatotoxicity? J Hepatol 2015;62:751–752.
- [21] Stephens C, Andrade RJ, Lucena MI. Mechanisms of drug-induced liver injury. Curr Opin Allergy Clin Immunol 2014;14:286–292.
- [22] Will Y, Dykens J. Mitochondrial toxicity assessment in industry-a decade of technology development and insight. Expert Opin Drug Metab Toxicol 2014;10:1061–1067.

- [23] Hughes JD, Blagg J, Price DA, Bailey S, Decrescenzo GA, Devraj RV, et al. Physiochemical drug properties associated with in vivo toxicological outcomes. Bioorg Med Chem Lett 2008;18:4872–4875.
- [24] Chen M, Bisgin H, Tong L, Hong H, Fang H, Borlak J, et al. Toward predictive models for drug-induced liver injury in humans: are we there yet? Biomark Med 2014;8:201–213.
- [25] Knowles SR, Uetrecht J, Shear NH. Idiosyncratic drug reactions: the reactive metabolite syndromes. Lancet 2000;356:1587–1591.
- [26] Pessayre D, Fromenty B, Berson A, Robin M-A, Lettéron P, Moreau R, et al. Central role of mitochondria in drug-induced liver injury. Drug Metab Rev 2012;44:34–87.
- [27] Faulkner L, Meng X, Park BK, Naisbitt DJ. The importance of hapten-protein complex formation in the development of drug allergy. Curr Opin Allergy Clin Immunol 2014;14:293–300.
- [28] Park B, Laverty H, Srivastava A, Antoine D, Naisbitt D, Williams D. Drug bioactivation and protein adduct formation in the pathogenesis of druginduced toxicity. Chem Biol Interact 2011;192:30–36.
- [29] Obach RS, Kalgutkar AS, Soglia JR, Zhao SX. Can in vitro metabolismdependent covalent binding data in liver microsomes distinguish hepatotoxic from nonhepatotoxic drugs? An analysis of 18 drugs with consideration of intrinsic clearance and daily dose. Chem Res Toxicol 2008;21:1814–1822.
- [30] Park BK, Boobis A, Clarke S, Goldring CEP, Jones D, Kenna JG, et al. Managing the challenge of chemically reactive metabolites in drug development. Nat Rev Drug Discov 2011;10:292–306.
- [31] Andrade RJ, Lucena MI, Martin-Vivaldi R, Fernandez MC, Nogueras F, Pelaez G, et al. Acute liver injury associated with the use of ebrotidine, a new H₂receptor antagonist. J Hepatol 1999;31:641–646.
- [32] Lucena MI, Andrade RJ, Rodrigo L, Salmerón J, Alvarez A, Lopez-Garrido M, et al. Trovafloxacin-induced acute hepatitis. Clin Infect Dis 2000;30:400–401.
- [33] Jaeschke H, McGill MR, Ramachandran A. Oxidant stress, mitochondria, and cell death mechanisms in drug-induced liver injury: lessons learned from acetaminophen hepatotoxicity. Drug Metab Rev 2012;44:88–106.
- [34] Xu JJ, Henstock PV, Dunn MC, Smith AR, Chabot JR, de Graaf D. Cellular imaging predictions of clinical drug-induced liver injury. Toxicol Sci 2008;105:97–105.
- [35] Boelsterli UA, Lim PL. Mitochondrial abnormalities A link to idiosyncratic drug hepatotoxicity? Toxicol Appl Pharmacol 2007;220:92–107.
- [36] Han D, Dara L, Win S, Than TA, Yuan L, Abbasi SQ, et al. Regulation of druginduced liver injury by signal transduction pathways: critical role of mitochondria. Trends Pharmacol Sci 2013;34:243–253.
- [37] Dykens JA, Will Y. The significance of mitochondrial toxicity testing in drug development. Drug Discov Today 2007;12:777–785.
- [38] Labbe G, Pessayre D, Fromenty B. Drug-induced liver injury through mitochondrial dysfunction: mechanisms and detection during preclinical safety studies. Fundam Clin Pharmacol 2008;22:335–353.
- [39] Chen M, Tung C-W, Shi Q, Guo L, Shi L, Fang H, et al. A testing strategy to predict risk for drug-induced liver injury in humans using high-content screen assays and the 'rule-of-two' model. Arch Toxicol 2014;88:1439–1449.
- [40] Porceddu M, Buron N, Roussel C, Labbe G, Fromenty B, Borgne-Sanchez A. Prediction of liver injury induced by chemicals in human with a multiparametric assay on isolated mouse liver mitochondria. Toxicol Sci 2012;129:332–345.
- [41] Pauli-Magnus C, Meier PJ. Hepatobiliary transporters and drug-induced cholestasis. Hepatology 2006;44:778–787.
- [42] Morgan RE, Trauner M, van Staden CJ, Lee PH, Ramachandran B, Eschenberg M, et al. Interference with bile salt export pump function is a susceptibility factor for human liver injury in drug development. Toxicol Sci 2010;118:485–500.
- [43] Aleo MD, Luo Y, Swiss R, Bonin PD, Potter DM, Will Y. Human drug-induced liver injury severity is highly associated to dual inhibition of liver mitochondrial function and bile salt export pump. Hepatology 2014;60:1015–1022.
- [44] Wu C-P, Hsieh C-H, Wu Y-S. The emergence of drug transporter-mediated multidrug resistance to cancer chemotherapy. Mol Pharm 2011;8:1996–2011.
- [45] Köck K, Ferslew BC, Netterberg I, Yang K, Urban TJ, Swaan PW, et al. Risk factors for development of cholestatic drug-induced liver injury: inhibition of hepatic basolateral bile acid transporters multidrug resistance-associated proteins 3 and 4. Drug Metab Dispos 2014;42:665–674.
- [46] Otani K, Kaneko S, Tasaki H, Fukushima Y. Hepatic injury caused by mianserin. BMJ 1989;299:519.
- [47] Seki N, Uematsu K, Shibakuki R, Eguchi K. Promising new treatment schedule for gefitinib responders after severe hepatotoxicity with daily administration. J Clin Oncol 2006;24:3213–3214.

- Drug Saf 2010;1:65–77.
 [49] Lucena MI, Andrade RJ, Kaplowitz N, García-Cortes M, Fernández MC, Romero-Gomez M, et al. Phenotypic characterization of idiosyncratic druginduced liver injury: the influence of age and sex. Hepatology 2009;49:2001–2009.
- [50] Chalasani N, Fontana RJ, Bonkovsky HL, Watkins PB, Davern T, Serrano J, et al. Causes, clinical features, and outcomes from a prospective study of drug-induced liver injury in the United States. Gastroenterology 2008;135:1924–1934.
- [51] Hunt CM, Yuen NA, Stirnadel-Farrant HA, Suzuki A. Age-related differences in reporting of drug-associated liver injury: data-mining of WHO Safety Report Database. Regul Toxicol Pharmacol 2014;70:519–526.
- [52] Dreifuss F, Santilli N, Langer D, Sweeney K, Moline K, Menander K. Valproic acid hepatic fatalities A retrospective review. Neurology 1987;37: 379–385.
- [53] Fountain FF, Tolley E, Chrisman CR, Self TH. Isoniazid hepatotoxicity associated with treatment of latent tuberculosis infection A 7-year evaluation from a public health tuberculosis clinic. Chest J 2005;128: 116–123.
- [54] Eichelbaum M, Ingelman-Sundberg M, Evans WE. Pharmacogenomics and individualized drug therapy. Annu Rev Med 2006;57:119–137.
- [55] Du H, Chen X, Fang Y, Yan O, Xu H, Li L, et al. Slow N-acetyltransferase 2 genotype contributes to anti-tuberculosis drug-induced hepatotoxicity: a meta-analysis. Mol Biol Rep 2013;40:3591–3596.
- [56] Urban TJ, Daly AK, Aithal GP. Genetic basis of drug-induced liver injury: present and future. Semin Liver Dis 2014;34:123–133.
- [57] Waxman DJ, Holloway MG. Sex differences in the expression of hepatic drug metabolizing enzymes. Mol Pharmacol 2009;76:215–228.
- [58] Hunt CM, Westerkam WR, Stave GM. Effect of age and gender on the activity of human hepatic CYP3A. Biochem Pharmacol 1992;44:275–283.
- [59] Morgan ET. Regulation of cytochromes P450 during inflammation and infection. Drug Metab Rev 1997;29:1129–1188.
- [60] Theken KN, Deng Y, Kannon MA, Miller TM, Poloyac SM, Lee CR. Activation of the acute inflammatory response alters cytochrome P450 expression and eicosanoid metabolism. Drug Metab Dispos 2011;39:22–29.
- [61] Seeff LB, Cuccherini BA, Zimmerman HJ, Adler E, Bendjamin SB. Acetaminophen hepatotoxicity in alcoholics A therapeutic misadventure. Ann Intern Med 1986;104:399–404.
- [62] Yu K, Geng X, Chen M, Zhang J, Wang B, Ilic K, et al. High daily dose and being a substrate of cytochrome P450 enzymes are two important predictors of drug-induced liver injury. Drug Metab Dispos 2014;42:744–750.
- [63] Huang Y-S, Su W-J, Huang Y-H, Chen C-Y, Chang F-Y, Lin H-C, et al. Genetic polymorphisms of manganese superoxide dismutase, NAD (P) H: quinone oxidoreductase, glutathione S-transferase M1 and T1, and the susceptibility to drug-induced liver injury. J Hepatol 2007;47:128–134.
- [64] Lucena MI, Andrade RJ, Martínez C, Ulzurrun E, García-Martín E, Borraz Y, et al. Glutathione S-transferase m1 and t1 null genotypes increase susceptibility to idiosyncratic drug-induced liver injury. Hepatology 2008;48:588–596.
- [65] Ng C-S, Hasnat A, Al Maruf A, Ahmed MU, Pirmohamed M, Day CP, et al. Nacetyltransferase 2 (NAT2) genotype as a risk factor for development of drug-induced liver injury relating to antituberculosis drug treatment in a mixed-ethnicity patient group. Eur J Clin Pharmacol 2014:1–8.
- [66] Lucena MI, García-Martín E, Andrade RJ, Martínez C, Stephens C, Ruiz JD, et al. Mitochondrial superoxide dismutase and glutathione peroxidase in idiosyncratic drug-induced liver injury. Hepatology 2010;52: 303–312.
- [67] Aruoma Ol. Free radicals, oxidative stress, and antioxidants in human health and disease. J Am Oil Chem Soc 1998;75:199–212.
- [68] Zawia NH, Lahiri DK, Cardozo-Pelaez F. Epigenetics, oxidative stress, and Alzheimer disease. Free Radic Biol Med 2009;46:1241–1249.
- [69] Finsterer J, Segall L. Drugs interfering with mitochondrial disorders. Drug Chem Toxicol 2010;33:138–151.
- [70] Kirchner H, Osler ME, Krook A, Zierath JR. Epigenetic flexibility in metabolic regulation: disease cause and prevention? Trends Cell Biol 2013;23:203–209.
- [71] Dai DF, Chiao YA, Marcinek DJ, Szeto HH, Rabinovitch PS. Mitochondrial oxidative stress in aging and healthspan. Longev Healthspan 2014;3:6.
- [72] Fromenty B. Drug-induced liver injury in obesity. J Hepatol 2013;58: 824–826.
- [73] Stewart JD, Horvath R, Baruffini E, Ferrero I, Bulst S, Watkins PB, et al. Polymerase γ Gene POLG determines the risk of sodium valproate-induced liver toxicity. Hepatology 2010;52:1791–1796.

[74] Felker D, Lynn A, Wang S, Johnson DE. Evidence for a potential protective effect of carnitine-pantothenic acid co-treatment on valproic acid-induced hepatotoxicity. Expert Rev Clin Pharmacol 2014;7:211–218.

JOURNAL OF HEPATOLOGY

- [75] Rodrigues AD, Lai Y, Cvijic ME, Elkin LL, Zvyaga T, Soars MG. Drug-induced perturbations of the bile acid pool, cholestasis, and hepatotoxicity: mechanistic considerations beyond the direct inhibition of the bile salt export pump. Drug Metab Dispos 2014;42:566–574.
- [76] de Vree JM, Jacquemin E, Sturm E, Cresteil D, Bosma PJ, Aten J, et al. Mutations in the MDR3 gene cause progressive familial intrahepatic cholestasis. Proc Natl Acad Sci U S A 1998;95:282–287.
- [77] Prieto J, García N, Martí-Climent JM, Peñuelas I, Richter JA, Medina JF. Assessment of biliary bicarbonate secretion in humans by positron emission tomography. Gastroenterology 1999;117:167–172.
- [78] Yoshikado T, Takada T, Yamamoto T, Yamaji H, Ito K, Santa T, et al. Itraconazole-induced cholestasis: involvement of the inhibition of bile canalicular phospholipid translocator MDR3/ABCB4. Mol Pharmacol 2011;79:241–250.
- [79] Daly AK, Donaldson PT, Bhatnagar P, Shen Y, Pe'er I, Floratos A, et al. HLA-B*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. Nat Genet 2009;41:816–819.
- [80] Cho J, Kim L, Li Z, Rose NR, Talor MV, Njoku DB. Sex bias in experimental immune-mediated, drug-induced liver injury in BALB/c mice: suggested roles for Tregs, estrogen, and IL-6. PLoS One 2013;8:e61186.
- [81] Toyoda Y, Miyashita T, Endo S, Tsuneyama K, Fukami T, Nakajima M, et al. Estradiol and progesterone modulate halothane-induced liver injury in mice. Toxicol Lett 2011;204:17–24.
- [82] Fontana RJ, Hayashi PH, Gu J, Reddy KR, Barnhart H, Watkins PB, et al. Idiosyncratic drug-induced liver injury is associated with substantial morbidity and mortality within 6 months from onset. Gastroenterology 2014;147:96–108.
- [83] Suzuki A, Yuen NA, Ilic Katarina, Hunt Christine M. Drug-induced liver injury (DILI) modulated by concomitant use of targeted drug classes: an analysis of 303 drugs associated with hepatotoxicity. Hepatology 2011;54:523A–524A.
- [84] Suzuki A, Yuen NA, Ilic Katarina, Hunt Christine M. Concomitant medications impact reporting frequency of drug-induced liver injury: data mining analysis using the WHO VigiBase[™] database. Hepatology 2011;54:523A.
- [85] Rizzardini M, Zappone M, Villa P, Gnocchi P, Sironi M, Diomede L, et al. Kupffer cell depletion partially prevents hepatic heme oxygenase 1 messenger RNA accumulation in systemic inflammation in mice: role of interleukin 1beta. Hepatology 1998;27:703–710.
- [86] Deng X, Stachlewitz RF, Liguori MJ, Blomme EA, Waring JF, Luyendyk JP, et al. Modest inflammation enhances diclofenac hepatotoxicity in rats: role of neutrophils and bacterial translocation. J Pharmacol Exp Ther 2006;319:1191–1199.
- [87] Shaw PJ, Hopfensperger MJ, Ganey PE, Roth RA. Lipopolysaccharide and trovafloxacin coexposure in mice causes idiosyncrasy-like liver injury dependent on tumor necrosis factor-alpha. Toxicol Sci 2007;100:259–266.
- [88] Wang LK, Wang LW, Li X, Han XQ, Gong ZJ. Ethyl pyruvate prevents inflammatory factors release and decreases intestinal permeability in rats with D-galactosamine-induced acute liver failure. Hepatobiliary Pancreat Dis Int 2013;12:180–188.
- [89] Nguyen GC, Sam J, Thuluvath PJ. Hepatitis C is a predictor of acute liver injury among hospitalizations for acetaminophen overdose in the United States: a nationwide analysis. Hepatology 2008;48:1336–1341.
- [90] Sulkowski MS. Drug-induced liver injury associated with antiretroviral therapy that includes HIV-1 protease inhibitors. Clin Infect Dis 2004;38: S90–S97.
- [91] Lomtadze N, Kupreishvili L, Salakaia A, Vashakidze S, Sharvadze L, Kempker RR, et al. Hepatitis C virus co-infection increases the risk of antituberculosis drug-induced hepatotoxicity among patients with pulmonary tuberculosis. PLoS One 2013;8:e83892.
- [92] Shu CC, Lee CH, Lee MC, Wang JY, Yu CJ, Lee LN. Hepatotoxicity due to firstline anti-tuberculosis drugs: a five-year experience in a Taiwan medical centre. Int J Tuberc Lung Dis 2013;17:934–939.
- [93] Suzuki A, Yuen N, Walsh J, Papay J, Hunt CM, Diehl AM. Co-medications that modulate liver injury and repair influence clinical outcome of acetaminophen-associated liver injury. Clin Gastroenterol Hepatol 2009;7:882–888.
- [94] Suzuki A, Watkins P, Kaplowitz N, Hunt C, Sanders C, Diehl A, et al. Comedication with adrenoreceptor antagonists is associated with lower meld scores at admission in patients with acetaminophen-induced acute liver failure. Gastroenterology 2009;136:A-810.
- [95] Robles-Diaz M, Lucena MI, Kaplowitz N, Stephens C, Medina-Caliz I, Gonzalez-Jimenez A, et al. Use of Hy's law and a new composite algorithm to predict acute liver failure in patients with drug-induced liver injury. Gastroenterology 2014;147:e105.

- [96] Shukla V, Cuenin C, Dubey N, Herceg Z. Loss of histone acetyltransferase cofactor transformation/transcription domain-associated protein impairs liver regeneration after toxic injury. Hepatology 2011;53:954–963.
- [97] Murata K, Hamada M, Sugimoto K, Nakano T. A novel mechanism for druginduced liver failure: inhibition of histone acetylation by hydralazine derivatives. J Hepatol 2007;46:322–329.
- [98] Brunaud L, Alberto JM, Ayav A, Gerard P, Namour F, Antunes L, et al. Effects of vitamin B12 and folate deficiencies on DNA methylation and carcinogenesis in rat liver. Clin Chem Lab Med 2003;41:1012–1019.
- [99] Zeisel SH. Dietary choline deficiency causes DNA strand breaks and alters epigenetic marks on DNA and histones. Mutat Res 2012;733:34–38.
- [100] Pooya S, Blaise S, Moreno Garcia M, Giudicelli J, Alberto JM, Gueant-Rodriguez RM, et al. Methyl donor deficiency impairs fatty acid oxidation through PGC-1alpha hypomethylation and decreased ER-alpha, ERR-alpha, and HNF-4alpha in the rat liver. J Hepatol 2012;57:344–351.
- [101] Halsted CH, Villanueva JA, Devlin AM, Niemela O, Parkkila S, Garrow TA, et al. Folate deficiency disturbs hepatic methionine metabolism and promotes liver injury in the ethanol-fed micropig. Proc Natl Acad Sci U S A 2002;99:10072–10077.
- [102] Villanueva JA, Esfandiari F, White ME, Devaraj S, French SW, Halsted CH. Sadenosylmethionine attenuates oxidative liver injury in micropigs fed ethanol with a folate-deficient diet. Alcohol Clin Exp Res 2007;31:1934–1943.
- [103] Schmucker DL, Sanchez H. Liver regeneration and aging: a current perspective. Curr Gerontol Geriatr Res 2011;2011:526379.
- [104] Brown GR, Persley K. Hepatitis A epidemic in the elderly. South Med J 2002;95:826–833.
- [105] Lewis J, Stine J. Review article: prescribing medications in patients with cirrhosis – A practical guide. Aliment Pharmacol Ther 2013;37:1132–1156.
- [106] Jog NR, Caricchio R. Differential regulation of cell death programs in males and females by Poly (ADP-Ribose) Polymerase-1 and 17beta estradiol. Cell Death Dis 2013;4:e758.
- [107] Ortona E, Matarrese P, Malorni W. Taking into account the gender issue in cell death studies. Cell Death Dis 2014;5:e1121.
- [108] Suzuki Ayako, Gu Jiezhun, Tillmann Hans, Bonkovsky Herbert, Fontana Robert, Kleiner David E. Association of gender and menopause with injury types and histological features of drug-induced liver injury. Gastroenterology 2014;146:S-1000–S-1001.
- [109] Ulzurrun E, Stephens C, Crespo E, Ruiz-Cabello F, Ruiz-Nuñez J, Saenz-López P, et al. Role of chemical structures and the 1331T> C bile salt export pump polymorphism in idiosyncratic drug-induced liver injury. Liver Int 2013;33:1378–1385.
- [110] Du L, Bayir H, Lai Y, Zhang X, Kochanek PM, Watkins SC, et al. Innate gender-based proclivity in response to cytotoxicity and programmed cell death pathway. J Biol Chem 2004;279:38563–38570.
- [111] Liang P, Lan F, Lee AS, Gong T, Sanchez-Freire V, Wang Y, et al. Drug screening using a library of human induced pluripotent stem cell-derived cardiomyocytes reveals disease specific patterns of cardiotoxicity. Circulation 2013;127:1677–1691.
- [112] Chen M, Shi L, Kelly R, Perkins R, Fang H, Tong W. Selecting a single model or combining multiple models for microarray-based classifier development? – A comparative analysis based on large and diverse datasets generated from the MAQC-II project. BMC Bioinformatics 2011;12:S3.
- [113] Bisgin H, Chen M, Wang Y, Kelly R, Fang H, Xu X, et al. A systems approach for analysis of high content screening assay data with topic modeling. BMC Bioinformatics 2013;14:1–10.
- [114] Ding Y, Chen M, Liu Z, Ding D, Ye Y, Zhang M, et al. AtBioNet An integrated network analysis tool for genomics and biomarker discovery. BMC Genomics 2012;13:325.
- [115] Hinton G, Osindero S, Teh Y-W. A fast learning algorithm for deep belief nets. Neural Comput 2006;18:1527–1554.
- [116] Chen M, Vijay V, Shi Q, Liu Z, Fang H, Tong W. FDA-approved drug labeling for the study of drug-induced liver injury. Drug Discov Today 2011;16:697–703.
- [117] Suzuki A, Andrade RJ, Bjornsson E, Lucena MI, Lee WM, Yuen NA, et al. Drugs associated with hepatotoxicity and their reporting frequency of liver adverse events in VigiBase: unified list based on international collaborative work. Drug Saf 2010;33:503–522.
- [118] Chen M, Borlak J, Tong W. Predicting idiosyncratic drug-induced liver injury-some recent advances. Expert Rev Gastroenterol Hepatol 2014;8:721–723.

- [119] Lammert C, Bjornsson E, Niklasson A, Chalasani N. Oral medications with significant hepatic metabolism at higher risk for hepatic adverse events. Hepatology 2010;51:615–620.
- [120] Cotreau MM, von Moltke LL, Greenblatt DJ. The influence of age and sex on the clearance of cytochrome P450 3A substrates. Clin Pharmacokinet 2005;44:33–60.
- [121] Chondrogianni N, Petropoulos I, Grimm S, Georgila K, Catalgol B, Friguet B, et al. Protein damage, repair and proteolysis. Mol Aspects Med 2014;35:1–71.
- [122] Ugarte N, Petropoulos I, Friguet B. Oxidized mitochondrial protein degradation and repair in aging and oxidative stress. Antioxid Redox Signal 2010;13:539–549.
- [123] Ruiz-Larrea M Begoña, Leal A Ma, Liza M, Lacort M, de Groot H. Antioxidant effects of estradiol and 2-hydroxyestradiol on iron-induced lipid peroxidation of rat liver microsomes. Steroids 1994;59:383–388.
- [124] Yang Z-X, Shen W, Sun H. Effects of nuclear receptor FXR on the regulation of liver lipid metabolism in patients with non-alcoholic fatty liver disease. Hepatol Int 2010;4:741–748.
- [125] Matsubara T, Li F, Gonzalez FJ. FXR signaling in the enterohepatic system. Mol Cell Endocrinol 2013;368:17–29.
- [126] Malik AF, Hoque R, Ouyang X, Ghani A, Hong E, Khan K, et al. Inflammasome components Asc and caspase-1 mediate biomaterial-induced inflammation and foreign body response. Proc Natl Acad Sci U S A 2011;108:20095–20100.
- [127] Begay CK, Gandolfi AJ. Late administration of COX-2 inhibitors minimize hepatic necrosis in chloroform induced liver injury. Toxicology 2003;185:79–87.
- [128] Lai I-R, Chang K-J, Tsai H-W, Chen C-F. Pharmacological preconditioning with simvastatin protects liver from ischemia-reperfusion injury by heme oxygenase-1 induction. Transplantation 2008;85:732–738.
- [129] Plaza-Diaz J, Gomez-Llorente C, Fontana L, Gil A. Modulation of immunity and inflammatory gene expression in the gut, in inflammatory diseases of the gut and in the liver by probiotics. World J Gastroenterol 2014;20:15632–15649.
- [130] Kloting N, Bluher M. Adipocyte dysfunction, inflammation and metabolic syndrome. Rev Endocr Metab Disord 2014;15:277–287.
- [131] Bhushan B, Borude P, Edwards G, Walesky C, Cleveland J, Li F, et al. Role of bile acids in liver injury and regeneration following acetaminophen overdose. Am J Pathol 2013;183:1518–1526.
- [132] Malkin CJ, Pugh PJ, Jones RD, Jones TH, Channer KS. Testosterone as a protective factor against atherosclerosis-immunomodulation and influence upon plaque development and stability. J Endocrinol 2003;178:373–380.
- [133] Cornacchia E, Golbus J, Maybaum J, Strahler J, Hanash S, Richardson B. Hydralazine and procainamide inhibit T cell DNA methylation and induce autoreactivity. J Immunol 1988;140:2197–2200.
- [134] Sacerdote P, Limiroli E, Gaspani L. Experimental evidence for immunomodulatory effects of opioids. Adv Exp Med Biol 2003;521:106–116.
- [135] Okamoto T, Iwata S, Ohnuma K, Dang N, Morimoto C. Histamine H1receptor antagonists with immunomodulating activities: potential use for modulating T helper type 1 (Th1)/Th2 cytokine imbalance and inflammatory responses in allergic diseases. Clin Exp Immunol 2009;157:27–34.
- [136] Kwak B, Mulhaupt F, Myit S, Mach F. Statins as a newly recognized type of immunomodulator. Nat Med 2000;6:1399–1402.
- [137] Bursch W, Karwan A, Mayer M, Dornetshuber J, Frohwein U, Schulte-Hermann R, et al. Cell death and autophagy: cytokines, drugs, and nutritional factors. Toxicology 2008;254:147–157.
- [138] Roberts SM, Harbison RD, Roth L, James RC. Methylphenidate-induced hepatotoxicity in mice and its potentiation by beta-adrenergic agonist drugs. Life Sci 1994;55:269–281.
- [139] Roberts SM, Harbison RD, Seng JE, James RC. Potentiation of carbon tetrachloride hepatotoxicity by phenylpropanolamine. Toxicol Appl Pharmacol 1991;111:175–188.
- [140] Fan M, Wang X, Xu G, Yan Q, Huang W. Bile acid signaling and liver regeneration. Biochim Biophys Acta 2015;1849(2):196–200.
- [141] Yokoyama Y, Nagino M, Nimura Y. Which gender is better positioned in the process of liver surgery? Male or female? Surg Today 2007;37:823–830.
- [142] Biondo-Simoes Mde L, Erdmann TR, Ioshii SO, Matias JE, Calixto HL, Schebelski DJ. The influence of estrogen on liver regeneration: an experimental study in rats. Acta Cir Bras 2009;24:3–6.

Review